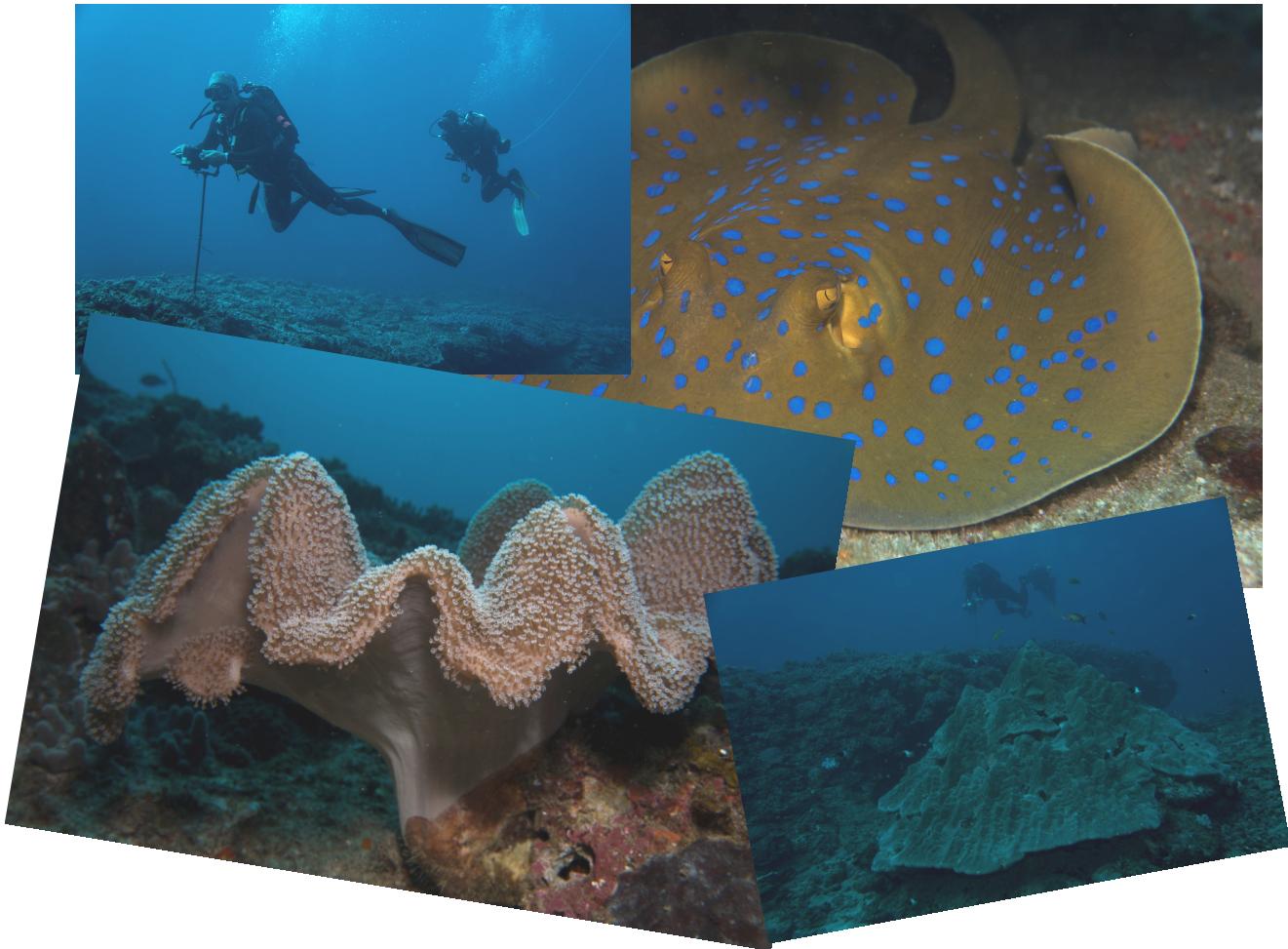




## REEF MONITORING IN THE PONTA DO OURO PARTIAL MARINE RESERVE: 2018



By  
Marcos A M Pereira, MSc.  
Cristina M M Louro, MApplSci.  
Raquel S Fernandes, MSc.



Maputo, May 2019



Centro Terra Viva - Estudos e Advocacia Ambiental



Reserva Marinha Parcial da Ponta do Ouro

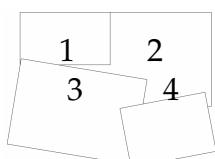
O Centro Terra Viva (CTV) e a Reserva Marinha Parcial da Ponta do Ouro (RMPPO), assinaram em Setembro de 2013 um Memorando de Entendimento (MdE) relativo a monitoria e investigação de espécies e ecossistemas da reserva. Este MdE foi posteriormente reforçado por um segundo MdE assinado entre o Fundo Nacional de Desenvolvimento Sustentável (FNDS), a Administração Nacional para as Áreas de Conservação (ANAC) e o (CTV), em Julho de 2017 que visa o desenvolvimento das actividades mencionadas acima em outras áreas de conservação marinhas, promovendo a sua protecção e conservação. A presente publicação resulta de actividades desenvolvidas no âmbito destes MdEs.

*Centro Terra Viva (CTV) and the Ponta do Ouro Partial Marine Reserve (POPMR) have established in September 2013 a Memorandum of Understanding (MoU) in order to develop several activities related to research and monitoring of species and ecosystems within the reserve. Te MoU was strengthened further by a second MoU signed between the National Fund for Sustainable Development (FNDS), the National Administration for Conservation Areas (ANAC) and CTV), in July 2017, in order to develop the activities mentioned above in other marine conservation areas, promoting their protection and conservation. The present publication is a result of activities undertaken under these MoUs.*

### Suggested citation:

Pereira, M.A.M., C.M.M. Louro & R.S. Fernandes (2019). Reef monitoring in the Ponta do Ouro Partial Marine Reserve: 2018, 15 pp. Maputo, CTV.

**Cover:** All photos by Jenny Strömvoll.



- 1 – Team conducting reef surveys (Kev's Ledge)
- 2 – Blue-spotted ribbontail ray, *Taeniura lymma* (Kev's Ledge)
- 3 – Close-up of a mushroom coral, *Sarcophyton* sp. (Kev's Ledge)
- 4 – A large coral head, *Porites* sp. (Kev's Ledge)

Maputo, May 2019

### Direitos Reservados

Direitos de autor aplicam-se a esta obra. Esta publicação seja por inteiro ou em partes, não poderá ser reproduzida independentemente do formato ou meio, seja electrónico, mecânico ou óptico, para qualquer propósito, sem a devida autorização expressa, por escrito, do Director Geral do Centro Terra Viva.

## ABSTRACT

This report presents data collected during the May/June 2018 reef benthic and fish monitoring surveys at the southern section of the Ponta do Ouro Partial Marine Reserve (POPMR). Data were collected on the five reefs selected using standardized methodology. Additionally, data on recreational divers' underwater behaviour were also collected. Coral cover was highest at Texas (43.1%) and lowest at Techobanine 2 with 24.4%. A low percentage of hard coral cover was found (<15% at all reefs), with soft coral dominating the reef biota (mainly *Lobophytum* and to a lesser degree *Sinularia*). Conversely, the percentage cover of rock/algae, rubble and sand was particularly high (ranging between 47 and 57%). The reef fish community was composed of typical small to medium reef fish, namely Chaetodontidae, Pomacentridae, Pomacanthidae and Labridae. The presence of large shoals of surgeon fishes (Acanthuridae), especially in the sanctuary reefs and of medium to large Serranidae and Scaridae species are good indicators of the health of these reefs. Overall, recreational SCUBA divers were relatively experienced with an average of 336.1 logged dives ( $SD= 101.5$ ). They made 5.6 ( $SD=6.9$ ) contacts with the substrata per 35 minute dive. No coral breakage by divers was observed, but abrasion and sediment resuspension were recorded.

## RESUMO

O presente relatório apresenta os dados da monitoria dos recifes, realizada em Maio/Junho de 2018, na secção sul da Reserva Marinha Parcial da Ponta do Ouro (RMPPO). Os dados foram colectados nos cinco recifes previamente seleccionados, usando metodologia standardizada. Adicionalmente, dados sobre o comportamento dos mergulhadores recreativos debaixo de água foram também colectados. A cobertura de coral vivo foi maior no recife Texas (43.1%) e menor em Techobanine 2 com 24.4%. Uma baixa percentagem de coral duro foi observada (<15% em todos recifes), sendo corais moles a categoria bentónica dominante (principalmente *Lobophytum* e de um modo menos significativo *Sinularia*). Por outro lado, a percentagem de cobertura de alga/rocha, calhau e areia foi particularmente alta (variando entre 47 e 57%). A comunidade ictiológica foi tipicamente composta por peixes de tamanho pequeno a médio, nomeadamente das famílias Chaetodontidae, Pomacentridae, Pomacanthidae e Labridae. A presença de grandes cardumes de cirurgiões (família Acanthuridae), especialmente nos recifes do sanctuário de Techobanine, e de garoupas (Serranidae) e papagaios (Scaridae) de tamanho médio a grande, constitui um bom indicador do estado de saúde destes recifes. De um modo geral, os mergulhadores recreativos eram relativamente experientes, com uma média de 336.1 mergulhos registados ( $DP=101.5$ ) e fizeram 5.6 ( $DP=6.9$ ) contactos com o substrato por mergulho de 35 min. Não foi observada nenhuma quebra de coral pelos mergulhadores, apesar de abrasão e resuspensão de sedimento terem sido observados.

## 1. INTRODUCTION

A reef monitoring program was established in 2011 (Pereira & Videira, 2011), after the proclamation of the Ponta do Ouro Partial Marine Reserve in 2009, building upon reef surveys that were conducted in 1996 (Robertson *et al.*, 1995) and 2002 (Pereira, 2003). This report presents the results of the fourth monitoring exercise, which included surveys of recreational SCUBA divers' underwater behaviour. Semi-quantitative reef fish surveys were also conducted and included in the compilation of species lists.

## 2. MATERIALS AND METHODS

### 2.1. Study Area

The Ponta do Ouro Partial Marine Reserve is located in southern Mozambique at the border with South Africa. A detailed description of the study reefs and general area is presented in Robertson *et al.* (1996) and Pereira (2003).

The five reefs previously chosen for the monitoring program at the southern section of the reserve were surveyed in May and June 2018: three reefs are located near Ponta Malongane (Creche–Cr, Kev's Ledge–KL and Texas–Tx) and two sites along the reef located off Mount Matonde, near Ponta Techobanine (Techo 1–Te1 and Techo 2–Te2). Table 1 and Figure 1 show the location of the reefs and GPS coordinates.

*Table 1. Location of selected reefs surveyed in 2016, at the POPMR.*

Reef code	Reef Name	GPS Coordinates (WGS84)		Observations
Cr	Creche	S26° 48.371	E32° 53.622	Offshore, subtidal patch reef. Depth: 10-14 m. Very high diving pressure.
KL	Kev's Ledge	S26° 46.673	E32° 54.268	Offshore, subtidal patch reef. Depth: 18-24 m. High diving pressure.
Te1	Techobanine 1	S26° 37.770	E32° 54.736	Offshore, subtidal patch reef. Depth: 16-20 m. Negligeable diving pressure.
Te2	Techobanine 2	S26° 37.806	E32° 54.873	Offshore, subtidal patch reef. Depth: 18-22 m. Negligeable diving pressure.
Tx	Texas	S26° 46.275	E32° 54.105	Offshore, subtidal patch reef. Depth: 12-18 m. Low diving pressure.

### 2.2. Reef Benthic Communities

The benthic communities were documented by a SCUBA diver using high-resolution, underwater digital imagery. The photographs were taken while swimming with the camera held at right angles to the reef face at a distance of 93 cm, the latter being regulated by a spacer bar attached to the camera housing. The distance between each photograph was 2-4 m, this being dictated by a pause in the camera recording system (Nikon Coolpix 4800). The area photographed in each photo-quadrat was approximately 0.3 m<sup>2</sup> and the distance between each photo-transect was at least 10 metres. The path of the transects was tracked using a floating GPS (Garmin eTrex). Annex 1 shows the coordinates of the transects at each reef. Each transect was composed of approximately 30 to 50 photo-quadrats. A total of 25 transects were completed, incorporating 941 photo-quadrats (Table 2).



**Figure 1.** Schematic map of the study area showing approximate location of the reefs surveyed (adapted from GoogleEarth). CR=Creche; KL=Kev's Ledge; Tx=Texas; Te1= Techobanine 1 and Te2=Techobanine 2.

**Table 2.** Sampling effort for reef benthos monitoring at the POPMR during the 2018 survey.

Reef	Date Surveyed	Transects	Photo-quadrats	Data points
Creche	31 May 2018	5	188	1 504
Kev's Ledge	30 May 2018	4	167	1 336
Techobanine 1	21 June 2018	5	172	1 376
Techobanine 2	21 June 2018	6	217	1 736
Texas	30 May 2018	5	197	1 576
Total		25	941	7 528

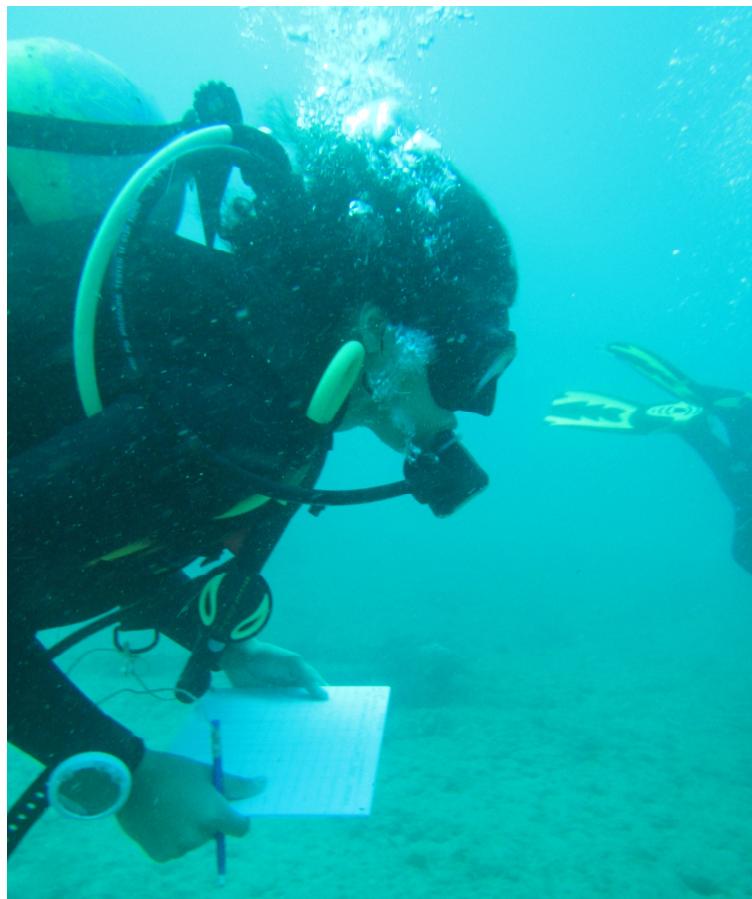
The data were extracted from the photo-quadrats using the point-intercept technique, where the images in JPEG format were analysed through the software CPCe 4.1 (Kohler & Gill, 2006). Eight randomly located points were superimposed on each image and the benthic category underneath each point identified to lowest possible taxonomic level. The morphological categories proposed by English *et al.* (1994) were used. A total of 7 528 random data points were analysed (Table 2).

### 2.3. Reef Fish Communities

The reef fish communities were monitored through direct observation and photography, using a Go-Pro camera. The monitoring consists in assessing fish abundance, through the indicative grouping of fish species as present (0 to 5 individuals), common (5 to 10 individuals) and abundant (10 or more individuals) and size classes through the registering of small (<15 cm), medium (15 – 30 cm) and large (> 30 cm). Fish species identification was made with the support of relevant literature (Lieske & Myers, 1999; King & Fraser, 2014).

### 2.4. Recreational SCUBA Divers' Underwater Behaviour

Underwater divers' behaviour surveys were carried out to assess and quantify the damage caused by divers to the benthic communities during two recreational dives in two shallow reefs, Doodles and Steve's Ledge. The methodology followed that described in Pereira (2003) and Pereira & Videira (2011), in order to ensure consistency and comparability of the data. A total of 11 divers were monitored, including two instructors.



*Figure 2. Diver collecting data on divers' behaviour. Photo: Raquel Fernandes.*

## 3. RESULTS

### 3.1. Reef Benthic Communities

At all reefs, the percentage cover of the general coral category was above 30%, except for Techobanine 2, with 25.6% (Table 3). Overall, the dominant category was rock and algae which, along with sand and rubble, covered between 47 to 57%. Fleshy macroalgae were recorded at all locations with Kev's Ledge ( $7.0 \pm 2.2\%$ ) and the two reefs at Techobanine (Techo 1 =  $8.7 \pm 2.7\%$  and Techo 2 =  $12.8 \pm 7.1$ ) showing the highest percentage cover. Another noteworthy aspect is the

significant presence of sponges (mainly encrusting) with 6.9% cover at Creche. While not very abundant, other invertebrates were seen commonly at all reefs and included both vasiform and encrusting sponges, sea stars (*Linkia* spp., *Culcita* spp.), sea cucumbers (*Stichopus* spp. and *Holothuria* spp.), and a vast array of molluscs. Not a single crown-of-thorns-starfish (CoTS; *Acanthaster mauritius*) or feeding scars were observed at any reef.

**Table 3.** Percentage cover ± SD of the major reef benthic categories at the POPMR during the 2016 survey.

Category	Creche	Kev's Ledge	Techo 1	Techo 2	Texas
Coral	38.5 ± 5.8	36.8 ± 5.7	32.0 ± 11.1	25.6 ± 23.1	43.2 ± 7.7
Macroalgae	1.4 ± 1.1	7.0 ± 2.2	8.7 ± 2.7	12.8 ± 7.1	3.7 ± 0.9
Coralline algae	0.0 ± 0.0	0.5 ± 0.3	0.1 ± 0.3	1.6 ± 3.2	0.1 ± 0.2
Molluscs	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	0.1 ± 0.2	0.0 ± 0.0
Ascidians	3.0 ± 1.5	3.6 ± 0.7	1.4 ± 1.3	0.6 ± 0.7	2.0 ± 1.1
Other invertebrates	7.3 ± 2.6	3.3 ± 1.0	1.7 ± 1.8	1.8 ± 1.0	3.2 ± 3.0
Sand, rock/algae, rubble	49.8 ± 4.3	46.8 ± 5.2	56.0 ± 8.8	57.4 ± 16.6	47.0 ± 5.7

In terms of corals, Texas had the highest total live coral coverage (43.1%) while Techobanine 2, with 24.4%, had the lowest cover (Table 4). Soft corals (mainly *Lobophytum* and to a lesser degree *Sinularia*; Annex 2) were the dominant biota (percentage cover ranged from around 14.1% in Techo 2 to 37.0% in Texas), with hard corals showing low cover at all locations, with the highest cover observed at Kev's Ledge (14.6% mainly encrusting *Montipora*) and Techo 2 (10.0% chiefly branching *Acropora*).

**Table 4.** Percentage cover ± SD of the major coral categories at the POPMR during the 2018 survey.

Category	Creche	Kev's Ledge	Techo 1	Techo 2	Texas
Branching hard coral	1.2 ± 0.6	1.9 ± 1.0	0.0 ± 0.0	8.5 ± 15.5	1.1 ± 0.8
Digitate hard coral	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0
Encrusting hard coral	1.0 ± 0.9	6.7 ± 1.4	0.5 ± 0.8	0.5 ± 0.6	1.8 ± 1.4
Foliose hard coral	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0
Free living coral	0.0 ± 0.0	0.2 ± 0.2	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0
Massive hard coral	2.9 ± 1.0	4.2 ± 1.8	3.50 ± 4.3	1.0 ± 0.9	2.2 ± 1.2
Submassive hard coral	0.1 ± 0.2	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	0.7 ± 1.5
Tabular hard coral	0.0 ± 0.0	1.1 ± 2.0	0.1 ± 0.2	0.0 ± 0.0	0.0 ± 0.0
Total hard coral	5.2 ± 1.9	14.6 ± 5.0	4.1 ± 4.1	10.0 ± 16.0	5.8 ± 1.8
Soft coral	32.8 ± 3.9	23.8 ± 7.5	27.5 ± 13.6	14.1 ± 11.0	37.0 ± 8.7
Fire coral	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0
Unidentified corals	0.4 ± 0.3	0.6 ± 0.2	0.1 ± 0.3	0.3 ± 0.3	0.3 ± 0.5
Total live coral	38.5 ± 5.8	38.0 ± 6.2	31.8 ± 11.1	24.4 ± 21.2	43.1 ± 7.8
Dead coral with algae	0.0 ± 0.0	0.2 ± 0.3	0.2 ± 0.2	1.2 ± 2.3	0.1 ± 0.1
Recently dead coral	0.0 ± 0.0	0.0 ± 0.0	0.1 ± 0.2	0.0 ± 0.0	0.0 ± 0.0

### 3.2. Reef Fish Communities

The reef fish species richness varied from 43 to 72 species, with the sanctuary reefs (i.e. Techobanine) presenting the least number of species (Table 5). This is probably the result of low coral cover and reef degradation which has been previously reported (Pereira, 2017). Nonetheless, continuous surveys need to be done to assess and monitor the fish communities on these reefs. The

cumulative species list for the POPMR is found in Annex 3. A total of 485 species (in 90 families) have been identified.

The fish communities of these reefs are represented by common coral reef species of small to medium size (e.g. Chaetodontidae, Labridae, Pomacentridae and Pomacanthidae). However, in all five reefs a common coral reef fish family Acanthuridae was observed in great numbers and with sizes ranging from small, medium to large. In the sanctuary reefs, medium to large individuals of *Acanthurus thompsoni* and *A. xanthopterus* were found in shoals. At Kev's Ledge, Creche and Texas large size individuals of parrotfishes (family Scaridae) were observed in abundant numbers, mainly *Scarus rubroviolaceus* e *S. scaber*. The rockcod family (Serranidae), was represented in all reefs. At Creche at least six species were identified, of which large individuals of *Varioula louti* and *Cephalopholis miniata* were also found in great numbers. At Kev's Ledge a shark was observed but identification to species level was not possible.

**Table 5.** Reef fish community diversity at the POPMR southern section reefs during the 2018 survey.

	Creche	Kev's Ledge	Techo 1	Techo 2	Texas
<b>Families</b>	18	21	21	14	18
<b>Species</b>	72	68	47	43	64
<i>Abundance (%)</i>					
<b>Present (0-5 ind.)</b>	33.3 (n=24)	64.7 (n=44)	51.1 (n=24)	55.8 (n=24)	62.5 (n=40)
<b>Common (5 – 10 ind.)</b>	12.5 (n=9)	13.0 (n=9)	27.7 (n=13)	18.6 (n=8)	14.1 (n=9)
<b>Abundant (&gt;10 ind.)</b>	54.2 (n=39)	22.0 (n=15)	21.3 (n=10)	25.6 (n=11)	23.4 (n=15)

In Techo 2 and Creche, two large marine turtles were identified, a hawksbill (*Eretmochelys imbricata*) and a green turtle (*Chelonia mydas*), respectively.

### 3.3. Recreational SCUBA Divers' Underwater Behaviour

The majority of the divers were males (54.5%). Only one diver (9.0%) had a camera. Overall, divers were relatively experienced with an average of 336.1 logged dives. Less experienced divers with only five dives were also monitored (Table 6).

**Table 6.** Summary of surveyed divers at the POPMR, 2018. SD=standard deviation.

Level of instruction	Nr of divers	Number of dives		
		Average ± SD	Min	Max
Advanced open water	4	72.0 ± 46.5	18	120
Dive Master	1	316.0		
Instructor	2	+1 500.0		
Junior open water	1	5.0		
Open water	3	29.3 ± 8.1	20	35
Total	11	336.1 ± 582.0	5	1 500

Table 7 presents the results of the surveys on the underwater behaviour of recreational SCUBA divers. On average, divers got in contact 5.6 times (SD = 6.9) with the substract, per 35 min dive. Only one diver with a camera was surveyed and made one contact with the substract. Intentional contacts were made with hands (66.7%) on other benthos (mostly rock; 33.3%), but the same

number of contacts were observed in hard coral and sand substract, and resulted mostly on abrasion (50.0%) and sediment resuspension (50.0%). As a result, no coral breakage was observed.

**Table 7.** Summary results of the underwater behavior of recreational SCUBA divers' surveys conducted from 2002 to 2018 at the POPMR.

Parameter	2002	2011	2014	2018
N Males	20	17	12	6
N Females	5	10	8	5
Average N dives ± SD	128.5 ± 230.5	202.0 ± 535.1	160.1 ± 342.5	336.1 ± 582.0
Average contacts/35 min dive ± SD	20.2 ± 38.4	13.7 ± 16.3	16.1 ± 22.4	5.6 ± 6.9
N divers with cameras (%)	2 (8)	8 (30)	3 (15)	1 (9)
N contacts: divers with cameras	54.3 ± 76.7	20.1 ± 14.7	22.2 ± 31.4	7
N contacts: divers without cameras	15.7 ± 30.1	11.1 ± 16.6	15.2 ± 19.8	4.9 ± 6.3
Contact by hand (%)	32.1	21.6	43.5	66.7
Contact by knee (%)	4.5	3.9	6.5	16.7
Contact by gear (%)	53.9	72.5	50.0	16.7
Contact by other (%)	7.5	2.0	0	0
Contacts on hard coral (%)	11.2	1.9	4.3	33.3
Contacts on soft coral (%)	14.2	9.6	2.2	0
Contacts on other benthos (%)	34.3	34.6	47.8	33.3
Contacts on sand (%)	40.3	53.8	45.7	33.3
Contacts with breakage (%)	2.2	2.0	0.0	0.0
Contacts with abrasion (%)	97.8 *	37.3	52.2	50.0
Contacts with sediment resuspension (%)	*	60.8	47.8	50.0

\* in 2002, "abrasion" included both abrasion and sediment resuspension.

## 4. CONCLUDING REMARKS

From the results of the 2018 monitoring survey, two aspects are worth mentioning:

### *Reef Recovery*

The coral communities seem to be recovering from the degradation previously reported by Pereira & Fernandes (2014), although at different paces. This is very encouraging and warrants consistent and systematic monitoring.

Despite the low number of divers sampled, the results of the surveys on divers' underwater behaviour suggest that awareness of good dive practice seems to be increasing. This is indicated by the average number of contacts divers made with the substract, in comparison to previous surveys (Pereira, 2003; Pereira & Videira, 2011; Pereira & Fernandes, 2014). Whilst it appears that the recreational SCUBA diving has minimal effect on the coral communities, management authorities are encouraged to continue to convey awareness about safe diving practices and to closely monitor the diving pressure on the reefs. Other factors, such as coral bleaching and the crown-of-thorns starfish (*Acanthaster mauritiensis*) should also merit attention. A close collaboration and liaison with the diving community should suffice in term of providing early warning of these factors.

In order to gain a wider and deeper understanding of the biodiversity, distribution and condition of the reefs within the reserve, reef assessments in other areas are recommended. These will provide

invaluable insight into the overall status of reefs in the POPMR, which will be especially valuable in view of the proclamation of the reserve as a UNESCO World Heritage Site.

#### *Fish Community*

The diversity and biomass of the reef fish communities indicates that these reefs are in good condition, especially based on the presence, in large numbers and sizes, of critical families such Acanthuridae, Scaridae and Serranidae in the majority of the reefs monitored. However, pelagic predators such as the king mackerel (*Scomberomorus commerson*), green jobfish (*Aprion virecens*) or even smaller specimens of tuna species were conspicuously absent. The surveys conducted were semi-quantitative in nature, and thus do not provide the true picture of the status of these fish communities. Therefore, rigorous monitoring of sport-fishing catches must continue in order to provide critical information on stocks as well as management recommendations.

## 5. ACKNOWLEDGMENTS

The Ponta do Ouro Partial Marine Reserve and Peace Parks Foundation are thanked for providing funding and logistical support. The National Administration for Conservation Areas (ANAC) authorized this study and provided financial support via the MozBio project. Centro Terra Viva, supplied a vehicle and contributed staff time. The following individuals and institutions provided invaluable support during field work: Miguel Gonçalves, Mark Eardly, Filimone Javane, Vicente Matsimbe, Álvaro Machaeie (POPMR), Elise da Fontoura and Jenny Strømvoll (Back to Basics Diving). Markus Joubert (Gozo Azul) and Sandy Probert (Oceana Diving), Petromoc, Toyota, Prodata and Satcom are also acknowledged for their support. Dr. Wanjiku Kiambo proof read the manuscript which helped improve its quality.

## 6. REFERENCES

- English, S., C. Wilkinson & V. Baker (eds) (1994). Survey manual for tropical marine resources. 2<sup>nd</sup> Edition, 368 pp. Townsville, Australian Institute of Marine Science.
- Haszprunar, G., C. Vogler & G. Wörheide (2017). Persistent gaps of knowledge for naming and distinguishing multiple species of crown-of-thorns-seastar in the *Acanthaster planci* species complex. *Diversity*, 9 (22): doi:10.3390/d9020022.
- King, D. & V. Fraser (2014). The reef guide – fishes, corals, nudibranchs & other invertebrates: East & south coasts of southern Africa. 360 pp. Cape Town, Struik Nature.
- Kohler, K. E. & S. M. Gill (2006). Coral Point Count with Excel extensions (CPCe): A Visual Basic program for the determination of coral and substrate coverage using random point count methodology. *Computers and Geosciences*, 32: 1259-1269.
- Lieske, E. & R. Myers (1999). Coral reef fishes – Caribbean, Indian Ocean, and Pacific Ocean including the Red Sea. 400 pp. Princeton, Princeton University Press.
- Pereira, M. A. M. (2003). Recreational SCUBA diving and reef conservation in southern Mozambique. MSc thesis. 109 pp. Durban, University of Natal.
- Pereira, M. A. M. (2017). Reef monitoring in the Ponta do Ouro Partial Marine Reserve 2016, 8 pp. Maputo, CTV.
- Pereira, M. A. M. & R. S. Fernandes (2014). Monitoring of reef communities in the Ponta do Ouro Partial Marine Reserve: 2014. 19 pp. Maputo, CTV.
- Pereira, M. A. M. & E. J. S. Videira (2011). Status assessment and monitoring of reef communities in the Ponta do Ouro Partial Marine Reserve. Technical Report 1: 2011 monitoring and status

- assessment. AICM Relatório não Publicado 16: 31 pp. Maputo, Associação para Investigação Costeira e Marinha.
- Robertson, W. D., M. H. Schleyer, P. J. Fielding, B. J. Tomalin, L. E. Beckley, S. T. Fennessy, R. P. van der Elst, S. Bandeira, A. Macia & D. Gove (1996). Inshore marine resources and associated opportunities for development of the coast of southern Mozambique: Ponta do Ouro to Cabo de Santa Maria. *South African Association for Marine Biological Research Unpublished Report* N° 130: 1–51. Durban, SAAMBR.
- Schleyer, M. (1998). Crown of thorns starfish in the Indian Ocean. *Reef Encounter*, 23: 25-27.

**ANNEX 1. GPS co-ordinates (WGS 84) of the photo-transects. Transects run either north or south depending on the prevailing current at the time of sampling.**

Reef (reef code)	Transect #	Start	End
Creche (Cr)	Tr1	26° 48.748 S ; 32° 53.646 E	26° 48.716 S ; 32° 53.640 E
	Tr2	26° 48.711 S ; 32° 53.636 E	26° 48.728 S ; 32° 53.630 E
	Tr3	26° 48.731 S ; 32° 53.629 E	26° 48.745 S ; 32° 53.621 E
	Tr4	26° 48.752 S ; 32° 53.622 E	26° 48.736 S ; 32° 53.628 E
	Tr5	26° 48.733 S ; 32° 53.628 E	26° 48.717 S ; 32° 53.632 E
Kev's Ledge (KL)	Tr1	26° 46.731 S ; 32° 54.243 E	26° 46.710 S ; 32° 54.246 E
	Tr2	26° 46.705 S ; 32° 54.246 E	26° 46.712 S ; 32° 54.252 E
	Tr3	26° 46.720 S ; 32° 54.254 E	26° 46.720 S ; 32° 54.259 E
	Tr4	26° 46.712 S ; 32° 54.257 E	26° 46.698 S ; 32° 54.241 E
Techobanine 1 (Te1)	Tr1	26° 37.775 S ; 32° 54.695 E	26° 37.759 S ; 32° 54.690 E
	Tr2	26° 37.753 S ; 32° 54.689 E	26° 37.739 S ; 32° 54.681 E
	Tr3	26° 37.732 S ; 32° 54.678 E	26° 37.721 S ; 32° 54.696 E
	Tr4	26° 37.720 S ; 32° 54.702 E	26° 37.699 S ; 32° 54.707 E
	Tr5	26° 37.693 S ; 32° 54.703 E	26° 37.675 S ; 32° 54.699 E
Techobanine 2 (Te2)	Tr1	26° 37.825 S ; 32° 54.838 E	26° 37.825 S ; 32° 54.824 E
	Tr2	26° 37.829 S ; 32° 54.822 E	26° 37.836 S ; 32° 54.788 E
	Tr3	26° 37.829 S ; 32° 54.779 E	26° 37.794 S ; 32° 54.779 E
	Tr4	26° 37.793 S ; 32° 54.777 E	26° 37.782 S ; 32° 54.774 E
	Tr5	26° 37.777 S ; 32° 54.776 E	26° 37.763 S ; 32° 54.761 E
	Tr6	26° 37.741 S ; 32° 54.755 E	26° 37.712 S ; 32° 54.742 E
Texas (Tx)	Tr1	26° 46.335 S ; 32° 54.024 E	26° 46.350 S ; 32° 54.033 E
	Tr2	26° 46.354 S ; 32° 54.036 E	26° 46.372 S ; 32° 54.048 E
	Tr3	26° 46.372 S ; 32° 54.055 E	26° 46.401 S ; 32° 54.060 E
	Tr4	26° 46.407 S ; 32° 54.070 E	26° 46.433 S ; 32° 54.066 E
	Tr5	26° 46.448 S ; 32° 54.063 E	26° 46.475 S ; 32° 54.062 E

**ANNEX 2. Percentage cover ± SD of coral genera identified at the within POPMR during the 2018 survey. % trans = percentage of the genera within the transect; % coral = percentage of the genera within corals only. SD = standard deviation.**

Genera	Creche				Kev's Ledge				Techo 1				Techo 2				Texas				
	% tran	SD	% coral	SD	% tran	SD	% coral	SD	% tran	SD	% coral	SD	% tran	SD	% coral	SD	% tran	SD	% coral	SD	
<i>Acanthastrea</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Acropora</i>	0.4	0.3	0.9	0.6	1.4	1.2	3.4	2.8	0.0	0.0	0.0	0.0	8.7	15.9	18.4	28.7	0.5	0.6	1.2	1.4	
<i>Alveopora</i>	0.1	0.2	0.2	0.4	0.1	0.1	0.2	0.3	0.0	0.0	0.0	0.0	0.3	0.6	0.6	1.0	1.0	1.3	2.7	3.5	
<i>Astreopora</i>	0.2	0.3	0.5	0.8	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.3	0.6	0.7	1.5	
<i>Cespitularia</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Cladiela</i>	1.2	1.3	2.8	2.9	0.0	0.0	0.0	0.0	0.2	0.2	0.8	1.1	0.0	0.0	0.0	0.0	0.0	0.3	0.7	0.8	1.8
<i>Dendronephthya</i>	0.1	0.2	0.2	0.4	1.8	1.5	4.9	4.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Diploastrea</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Dipsastraea</i>	0.3	0.3	0.7	0.7	1.6	1.1	4.2	2.4	0.1	0.2	0.4	1.0	0.1	0.1	3.3	8.2	0.5	0.3	1.2	0.8	
<i>Echinopora</i>	0.6	1.0	1.3	2.1	1.0	1.1	3.0	3.7	0.0	0.0	0.0	0.0	0.1	0.2	0.4	0.9	0.6	0.9	1.3	1.8	
<i>Favites</i>	1.5	0.6	3.8	1.6	1.5	0.6	3.7	1.1	0.5	0.9	2.3	4.7	0.1	0.2	0.3	0.5	0.4	0.3	0.9	0.8	
<i>Fungiid</i>	0.0	0.0	0.0	0.0	0.2	0.2	0.4	0.5	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.1	0.2	0.2	0.5	
<i>Galaxea</i>	0.3	0.2	0.9	0.6	0.1	0.1	0.2	0.3	0.2	0.3	1.1	1.6	0.0	0.0	0.0	0.0	0.1	0.2	0.2	0.5	
<i>Gardineroseris</i>	0.2	0.4	0.5	1.1	0.2	0.3	0.6	0.8	0.0	0.0	0.0	0.0	0.1	0.1	3.3	8.2	0.0	0.0	0.0	0.0	
<i>Goniastrea</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.1	0.1	0.2	0.4	0.0	0.0	0.0	0.0	
<i>Goniopora</i>	0.1	0.2	0.3	0.6	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.1	0.1	0.2	0.4	
<i>Gorgonian</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Hydnophora</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Leptoseris</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Lobophyllia</i>	0.0	0.0	0.0	0.0	0.1	0.2	0.3	0.5	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Lobophytum</i>	28.6	2.2	75.1	8.7	15.9	3.8	41.1	6.0	19.2	12.6	56.1	20.6	8.7	7.8	47.3	31.9	29.9	6.2	69.0	5.7	
<i>Merulina</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Montipora</i>	0.3	0.4	0.9	1.0	5.4	0.7	14.5	3.9	0.5	0.8	2.2	3.5	0.1	0.3	0.5	0.9	1.2	0.8	2.7	2.1	
<i>Mycedium</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Nephthiid</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Oxypora</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Pachyseris</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Pavona</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Platygyra</i>	0.4	0.5	1.1	1.2	0.5	0.4	1.1	1.0	1.4	2.1	4.9	7.2	0.4	0.6	4.6	7.9	0.4	0.5	1.0	1.4	
<i>Plesiastrea</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
<i>Pocillopora</i>	0.7	0.6	1.8	1.4	0.7	0.8	2.1	2.8	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.5	0.5	1.3	1.2	
<i>Porites</i>	0.1	0.1	0.2	0.4	0.1	0.2	0.2	0.4	1.3	2.6	4.5	8.4	0.0	0.0	0.0	0.0	0.1	0.3	0.3	0.6	

## ANNEX 2. Cont.

Genera	Creche				Kev's Ledge				Techo 1				Techo 2				Texas			
	% tran	SD	% coral	SD	% tran	SD	% coral	SD	% tran	SD	% coral	SD	% tran	SD	% coral	SD	% tran	SD	% coral	SD
<i>Rhytisma</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Rumphella</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.1	0.2	0.2	0.4	0.6	1.0	2.2	3.5	0.0	0.0	0.0	0.0
<i>Sarcophyton</i>	0.4	0.5	1.1	1.6	0.6	0.6	1.5	1.4	1.9	1.2	7.0	5.1	0.4	0.6	1.4	2.3	1.4	0.6	3.2	1.0
<i>Seriatopora</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Simularia</i>	2.6	2.6	6.2	5.2	5.3	3.6	13.1	8.1	6.1	3.4	18.3	5.5	4.3	4.3	13.9	15.9	5.2	2.8	11.7	4.2
<i>Stylophora</i>	0.1	0.2	0.3	0.4	0.1	0.2	0.3	0.5	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.1	0.2	0.2	0.5
<i>Tubipora</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.1	0.2	0.2	0.5	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Turbinaria</i>	0.0	0.0	0.0	0.0	1.0	2.0	2.5	4.9	0.1	0.2	0.2	0.4	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Xeniid</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Bleached coral	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Disease	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0

**ANNEX 3. Cumulative checklist of reef and litoral fish species identified at the POPMR. The species list was compiled from Pereira et al. (2004), Floros (2010), Pereira & Videira (2014) and Pereira (2016). \* = species observed in 2018; (?) = to be confirmed.**

**Acanthuridae**

*Acanthurus auranticavus*  
*Acanthurus blochii\**  
*Acanthurus dussumieri\**  
*Acanthurus leucosternon\**  
*Acanthurus lineatus*  
*Acanthurus mata*  
*Acanthurus nigrofasciatus\**  
*Acanthurus tennentii\**  
*Acanthurus thompsoni\**  
*Acanthurus triostegus*  
*Acanthurus xanthopterus\**  
*Ctenochaetus binotatus\**  
*Ctenochaetus striatus\**  
*Ctenochaetus truncatus\**  
*Naso brachycentron*  
*Naso brevirostris*  
*Naso elegans\**  
*Naso fageni*  
*Naso hexacanthus\**  
*Naso unicornis*  
*Paracanthurus hepatus*  
*Zebrassoma gemmatum\**  
*Zebrassoma scopas\**

**Antennariidae**

*Antennatus coccineus*  
*Antennatus nummifer*

**Apogonidae**

*Apogon coccineus*  
*Apogon semiornatus*  
*Apogonichthys ocellatus*  
*Cheilodipterus artus*  
*Cheilodipterus macrodon*  
*Gymnapogon africanus*  
*Neamia octospina*  
*Ostorhinchus angustatus*  
*Ostorhinchus apogonides*  
*Ostorhinchus aureus*  
*Ostorhinchus taeniophorus*  
*Pristiapogon fraenatus*  
*Pristiapogon kallopterus*  
*Siphamia mossambica*

**Atherinidae**

*Atherinomorus lacunosus*

**Aulostomidae**

*Aulostomos chinensis*

**Balistidae**

*Aluterus scriptus*

*Balistapus undulatus\**  
*Balistoides conspicillum\**  
*Balistoides viridescens\**  
*Melichthys indicus\**  
*Melichthys niger*  
*Odonus niger\**  
*Pseudobalistes fuscus*  
*Rhinechanthus aculeatus\**  
*Sufflamen bursa*  
*Sufflamen chrysopterum\**  
*Sufflamen fraenatum*

**Blenniidae**

*Alloblennius parvus*  
*Aspidontus dussumieri\**  
*Aspidontus tractus*  
*Cirripectes castaneus*  
*Ecsenius midas*  
*Ecsenius nalolo*  
*Exallias brevis*  
*Hirculops cornifer*  
*Blenniella cyanostigma*  
*Istiblennius dussumieri*  
*Istiblennius edentulus*  
*Pterulixia kosiensis*  
*Plagiotremus rhinorhynchos*  
*Plagiotremus tapeinosoma*  
*Scartella emarginata*

**Bothidae**

*Bothus mancus*  
*Dinematicthys sp.*  
*Engyproposon sp.*

**Caesionidae**

*Caesio caeruleaurea*  
*Caesio lunaris*  
*Caesio teres*  
*Caesio xanthonota\**  
*Caseio xanthalythos?*  
*Pteroacesio tile*

**Callionymidae**

*Callionymus marleyi*  
*Synchiropus postulus*  
*Synchiropus stellatus*

**Caracanthidae**

*Caracanthus madagascariensis*  
*Caracanthus unipinna*

**Carangidae**

*Alectis ciliaris*

**Alectis indica**

*Alepes djedaba*  
*Carangoides oaeruleopinnatus*  
*Carangoides ferdau*  
*Carangoides fulvoguttatus*  
*Caranx heberi*  
*Caranx ignobilis*  
*Caranx melampygus*  
*Caranx papuensis*  
*Caranx sexfasciatus*  
*Decapterus macarellus*  
*Gnathanodon speciosus*  
*Pseudocaranx dentex*  
*Scomberoides lysan*  
*Seriolina nigrofasciata*  
*Trachinotus botla*

**Carcharhinidae**

*Carcharhinus amblyrhynchos*  
*Carcharhinus leucas*  
*Galeocerdo cuvier*  
*Triaenodon obesus\**

**Chaetodontidae**

*Chaetodon auriga\**  
*Chaetodon blackburnii\**  
*Chaetodon dolosus*  
*Chaetodon guttatissimus\**  
*Chaetodon interruptus\**  
*Chaetodon kleinii\**  
*Chaetodon lunula\**  
*Chaetodon madagaskariensis\**  
*Chaetodon meyeri\**  
*Chaetodon trifascialis\**  
*Chaetodon trifasciatus*  
*Chaetodon vagabundus*  
*Chaetodon zanzibariensis?*  
*Forcipiger flavissimus\**  
*Hemitaurichthys zoster\**  
*Heniochus acuminatus*  
*Heniochus diphreutes*  
*Heniochus monoceros*

**Cirrhitidae**

*Amblycirrhitus bimacula*  
*Cirrhitichthys oxycephalus\**  
*Paracirrhites arcatus\**  
*Paracirrhites forsteri\**

**Clinidae**

*Pavoclinus graminis*  
*Pavoclinus laurentii*

<b>Congridae</b>	<i>Heteroleotris tentaculata</i>	<i>Anampsese lineatus</i>
<i>Conger cinereus</i>	<i>Heteroleotris zonata</i>	<i>Anampsese meleagrides*</i>
<b>Congrogadidae</b>	<i>Istigobius decoratus</i>	<i>Anampsese twistii*</i>
<i>Halimuraena shakai</i>	<i>Nemateleotris magnifica</i>	<i>Bodianus anthiooides</i>
<b>Coracinidae</b>	<i>Pleurosicya mossambica</i>	<i>Bodianus atrolumbus*</i>
<i>Dichistius multifasciatys</i>	<i>Priolepis cincta</i>	<i>Bodianus axillaris</i>
<b>Creediidae</b>	<i>Ptereleotris eteroptera</i>	<i>Bodianus bilunulatus*</i>
<i>Apodocreedia vanderhorsti</i>	<i>Ptereleotris evides</i>	<i>Bodianus diana*</i>
<i>Limnichthys nitidus</i>	<i>Trimma macrophthalmum</i>	<i>Bodianus perditio</i>
<b>Cynoglossidae</b>	<i>Valenciennea strigata</i>	<i>Bodianus trilineatus</i>
<i>Cynoglossus sp.</i>		<i>Calotomus carolinus</i>
<i>Parapaglusia bilineata</i>		<i>Cheilinus trilobatus</i>
<b>Dasyatidae</b>		<i>Cheilio inermis</i>
<i>Maculabatis gerrardi</i>		<i>Chlorurus atrilunula</i>
<i>Neotrygon kuhlii</i>		<i>Chlorurus cyanescens</i>
<i>Taeniura lymma</i>		<i>Cirrhilabrus exquisitus</i>
<i>Taeniura meyeni</i>		<i>Coris aygula</i>
<i>Urogyminus asperrimus</i>		<i>Coris caudimacula</i>
<b>Dinopercidae</b>		<i>Coris cuvieri</i>
<i>Dinopercia petersi</i>		<i>Coris cuvieri*</i>
<b>Diodontidae</b>		<i>Coris formosa*</i>
<i>Diodon hystric</i>		<i>Gomphosus caeruleus*</i>
<i>Diodon liturosus</i>		<i>Halichoeres cosmetus</i>
<b>Echeneidae</b>		<i>Halichoeres hortulanus*</i>
<i>Echeneis naucrates</i>		<i>Halichoeres iridis*</i>
<b>Ephippidae</b>		<i>Halichoeres nebulosus</i>
<i>Platax teira</i>		<i>Halichoeres scapularis*(?)</i>
<i>Tripteronodon orbis</i>		<i>Hemigymnus fasciatus</i>
<b>Exocoetidae</b>		<i>Hologymnosus annulatus</i>
<i>Exocoetidae sp.</i>		<i>Hologymnosus doliatu</i>
<i>Fistularia commersonii</i>		<i>Iniistius pavo</i>
<i>Fistularia petimba</i>		<i>Labroides bicolor</i>
<i>Fistularidae</i>		<i>Labroides dimidiatus*</i>
<b>Gerreidae</b>		<i>Labropsis xanthonota</i>
<i>Gerres longirostris</i>		<i>Macropharyngodon bipartitus*</i>
<b>Gobiesocidae</b>		<i>Macropharyngodon cyanoguttatus</i>
<i>Lepidichthys coccinotaenia</i>		<i>Macropharyngodon vivienae*</i>
<b>Gobiidae</b>		<i>Novaculichthys taeniourus</i>
<i>Callogobius sclateri</i>		<i>Oxycheilinus bimaculatus</i>
<i>Eviota prasina</i>		<i>Oxycheilinus diogramma</i>
<i>Fusigobius duospillus</i>		<i>Pseudocheilinus evanidu</i>
<i>Fusigobius longispinus</i>		<i>Pseudocheilinus hexataenia</i>
<i>Gnatholepis sp.</i>		<i>Pseudodax molluccanus*</i>
<i>Gobiodon rivulatus</i>		<i>Pseudojuloides cerasinus</i>
<b>Helcogrammatidae</b>		<i>Stethojulis albovittata</i>
<b>Hemirhamphidae</b>		<i>Stethojulis interrupta</i>
<i>Hemiramphus affinis</i>		<i>Sthethojulis strigiventer</i>
<b>Holocentridae</b>		<i>Thalassoma amblycephalum</i>
<i>Myripristis berndti</i>		<i>Thalassoma genivittatum</i>
<i>Myripristis botche</i>		<i>Thalassoma hebraicum</i>
<i>Myripristis kuntee</i>		<i>Thalassoma lunare*</i>
<i>Myripristis murdjan*</i>		<i>Thalassoma lutescens</i>
<i>Myripristis vittata*</i>		<i>Thalassoma purpureum</i>
<i>Neoniphon argenteus</i>		<i>Thalassoma trilobatum</i>
<i>Neoniphon sammara</i>		<b>Scaridae</b>
<i>Sargocentrom ittodai</i>		<i>Calotomus carlinus*(?)</i>
<i>Sargocentron caudimaculatum*</i>		
<i>Sargocentron diadema*</i>		
<b>Istiophoridae</b>		
<i>Istiophorus platypterus</i>		
<i>Istiompax indica</i>		
<b>Khuliidae</b>		
<i>Khulia mugil</i>		
<b>Kraemeriidae</b>		
<i>Kraemeria samoensis</i>		
<b>Kyphosidae</b>		
<i>Kyphosus bigibbus</i>		
<b>Labridae</b>		
<i>Anampsese caeruleopunctatus</i>		

<i>Scarus frenatus</i>	<i>Crenimugil buchanani</i>	<b>Orectolobidae</b>
<i>Scarus ghobban</i>		<i>Stegostoma fasciatum</i>
<i>Scarus rubroviolaceus*</i>		
<i>Scarus scaber?(?)</i>		<b>Ostraciidae</b>
<i>Scarus tricolor*</i>		<i>Ostracion cubicus</i>
<i>Scarus viridifucatus</i>		<i>Ostracion meleagris</i>
<b>Lethrinidae</b>		
<i>Gnathodentex aureolineatus</i>		<b>Pempheridae</b>
<i>Gymnocranius grandoculis*</i>		<i>Parapriacanthus ransonneti*</i>
<i>Gymnocranius griseus</i>		<i>Pempheris adusta</i>
<i>Lethrinus crocineus</i>		<i>Pempheris schwenkii*</i>
<i>Lethrinus harak</i>		
<i>Lethrinus lentjan</i>		<b>Pinguipedidae</b>
<i>Lethrinus microdon*</i>		<i>Parapercis hexophtalma</i>
<i>Lethrinus nebulosus*</i>		<i>Parapercis punctulata</i>
<i>Lethrinus variegatus</i>		<i>Parapercis robinsoni</i>
<i>Monotaxis grandoculis</i>		<i>Parapercis sp.</i>
<b>Lutjanidae</b>		
<i>Aphareus furca</i>		<b>Platycephalidae</b>
<i>Aphareus rutilans</i>		<i>Onigocia oligolepis</i>
<i>Aprion virescens*</i>		<i>Thysanophrys chiltonae</i>
<i>Lutjanus argentinimaculatus</i>		<i>Sunagocia otaitensis</i>
<i>Lutjanus bohar*</i>		
<i>Lutjanus fulviflamma</i>		<b>Pleuronectidae</b>
<i>Lutjanus gibbus*</i>		<i>Samariscus triocellatus</i>
<i>Lutjanus kasmira*</i>		
<i>Lutjanus lutjanus*</i>		<b>Plotosidae</b>
<i>Lutjanus monostigma</i>		<i>Plotosus lineatus</i>
<i>Lutjanus rivulatus</i>		
<i>Lutjanus russellii</i>		<b>Polynemidae</b>
<i>Macolor niger*</i>		<i>Polydactylus plebeius</i>
<i>Paracaesio sordida*</i>		
<i>Paracaesio xanthura*</i>		<b>Pomacanthidae</b>
<b>Malacanthidae</b>		<i>Apolemichthys kingi</i>
<i>Malacanthus brevirostris</i>		<i>Apolemichthys trimaculatus*</i>
<i>Malacanthus latovittatus</i>		<i>Centropyge acanthops</i>
<b>Mobulidae</b>		<i>Centropyge bispinosa</i>
<i>Mobula birostris</i>		<i>Centropyge multispinis*</i>
<b>Monacanthidae</b>		<i>Genicanthus caudovittatus</i>
<i>Cantherines dumerilii</i>		<i>Pomacanthus imperator*</i>
<i>Cantherines fronticinctus</i>		<i>Pomacanthus rhomboides*</i>
<i>Cantherines pardalis</i>		<i>Pomacanthus semicirculatus*</i>
<i>Paralutereres prionurus</i>		<i>Pygoplites diacanthus</i>
<i>Pervagor janthinosoma</i>		
<i>Stephanolepis auratus</i>		<b>Pomacentridae</b>
<b>Monodactylidae</b>		<i>Abudefduf natalensis*</i>
<i>Monodactylus argenteus</i>		<i>Abudefduf notatus</i>
<b>Mugilidae</b>		<i>Abudefduf sexfasciatus</i>
<i>Planiliza macrolepis</i>		<i>Abudefduf sordidus</i>
<i>Mugil cephalus</i>		<i>Abudefduf sparoides</i>
		<i>Abudefduf vaigiensis</i>
		<i>Amphiprion akallopis</i> *
		<i>Amphiprion allardi*</i>
		<i>Chromis dasypenrys*</i>
		<i>Chromis fieldi*</i>
		<i>Chromis lepidolepis</i>
		<i>Chromis nigrura*</i>

<i>Chromis opercularis</i>	<i>Scorpaenodes parvipinnis</i>	<i>Rhabdosargus holubi</i>
<i>Chromis weberi*</i>	<i>Scorpaenodes varipinnis</i>	<i>Rhabdosargus sarba</i>
<i>Chrysiptera unimaculata</i>	<i>Scorpaenopsis brevifrons</i>	<i>Rhabdosargus thorpei</i>
<i>Dascyllus aruanus</i>	<i>Scorpaenopsis oxycephala</i>	
<i>Dascyllus carneus</i>	<i>Scorpaenopsis venosa</i>	
<i>Dascyllus trimaculatus*</i>	<i>Sebastapistes cyanostigma</i>	<b>Sphyraenidae</b>
<i>Neopomacentrus cyanomos</i>	<i>Sebastapistes mauritiana</i>	<i>Sphyraena jello</i>
<i>Plectroglyphidodon dickii</i>	<i>Sebastapistes strongia</i>	
<i>Plectroglyphidodon jhonstonianus</i>	<i>Taenianotus triacanthus</i>	
<i>Plectroglyphidodon lacrymatus</i>		<b>Sphyrnidae</b>
<i>Plectroglyphidodon leucozonus</i>		<i>Sphyrna sp.</i>
<i>Pomacentrus caeruleus*</i>		
<i>Pomacentrus pavo</i>		<b>Syngnathidae</b>
<i>Pomacentrus trichourus</i>		<i>Doryrhamphus excisus excisus</i>
 <b>Pomatomidae</b>		
<i>Pomatomus saltatrix</i>		<b>Synodontidae</b>
 <b>Priacanthidae</b>		<i>Saurida gracilis</i>
<i>Heteropriacanthus cruentatus</i>		<i>Synodus binotatus</i>
<i>Priacanthus hamrur*</i>		<i>Synodus dermatogenys*</i>
 <b>Pseudochromidae</b>		<i>Synodus jaculum</i>
<i>Chlidichthys johnvoelckeri</i>		<i>Synodus variegatus</i>
<i>Pseudochromis dutoiti</i>		
<i>Pseudochromis melas</i>		 <b>Teraponidae</b>
<i>Pseudochromis natalensis</i>		<i>Terapon jarbua</i>
 <b>Pseudogrammidae</b>		
<i>Pseudogramma polyacantha</i>		 <b>Tetraodontidae</b>
 <b>Ptereleotridae</b>		<i>Amblyrhynchotes honckenii</i>
<i>Ptereleotris sp.</i>		<i>Arothron hispidus</i>
 <b>Rhincodontidae</b>		<i>Arothron meleagris</i>
<i>Rhincodon typus</i>		<i>Arothron nigropunctatus*</i>
 <b>Rhinobatidae</b>		<i>Arothron stellatus</i>
<i>Rhynchosciurus djiddensis</i>		<i>Canthigaster ambionensis</i>
 <b>Sciaenidae</b>		<i>Canthigaster bennetti</i>
<i>Argyrosomus japonicus</i>		<i>Canthigaster cyanospilota</i>
<i>Umbrina canariensis</i>		<i>Canthigaster janthinoptera</i>
<i>Umbrina robinsoni</i>		<i>Canthigaster smithae</i>
 <b>Scombridae</b>		<i>Canthigaster valentini</i>
<i>Acanthocybium solandri</i>		
<i>Sarda orientalis</i>		 <b>Torpedinidae</b>
<i>Scomberomorus commerson*</i>		<i>Torpedo sinuspersici</i>
<i>Thunnus albacares</i>		
 <b>Scorpaenidae</b>		 <b>Trichonotidae</b>
<i>Parascorpaena mcdamisi</i>		<i>Trichonotus marleyi</i>
<i>Parascorpaena mossambica</i>		<i>Enneapterygius abeli</i>
<i>Pterois miles</i>		<i>Enneapterygius elegans</i>
<i>Pterois mombasae</i>		<i>Enneapterygius pusillus</i>
<i>Scorpaenodes kelloggi</i>		<i>Enneapterygius ventermaculatus</i>
		<i>Helcogramma fuscopinna</i>
		<i>Helcogramma obtusirostris</i>
		 <b>Zanclidae</b>
		<i>Zanclus cornutus*</i>